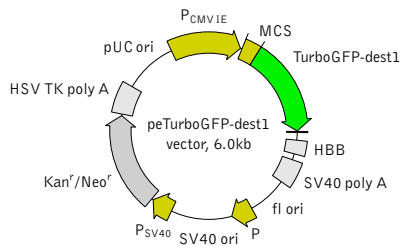


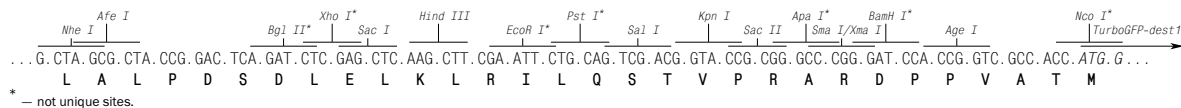
peTurboGFP-dest1 vector

The vector sequence has been compiled using the information from sequence databases, published literature, and other sources, together with partial sequences obtained by Evrogen. This vector has not been completely sequenced.



For vector sequence, please visit our Web site at <http://www.evrogen.com/products/vectors.shtml>

Multiple cloning site (MCS)



Location of features

P_{CMV IE}: 1-589
 Enhancer region: 59-465
 TATA box: 554-560
 Transcription start point: 583
 MCS: 591-671
 TurboGFP-dest1
 Kozak consensus translation initiation site: 672-682
 Start codon (ATG): 679-681
 Last amino acid in TurboGFP: 1372-1374
 Amino acid residues of mouse ornithine decarboxylase (MODC) PEST sequence: 1390-1509
 Stop codon: 1510-1512
 Fragment of human beta globin (HBB) gene
 Last 35 bp of HBB exon 2: 1521-1555
 HBB intron 2: 1556-2406
 First 233 bp of HBB exon 3: 2407-2639
 SV40 early mRNA polyadenylation signal
 Polyadenylation signals: 2781-2786 & 2810-2815
 mRNA 3' ends: 2819 & 2831
 f1 single-strand DNA origin: 2878-3333
 Bacterial promoter for expression of Kan^r gene
 -35 region: 3395-3400; -10 region: 3418-3423
 Transcription start point: 3430
 SV40 origin of replication: 3674-3809
 SV40 early promoter
 Enhancer (72-bp tandem repeats): 3507-3578 & 3579-3650
 21-bp repeats: 3654-3674, 3675-3695 & 3697-3717
 Early promoter element: 3730-3736
 Major transcription start points: 3726, 3764, 3770 & 3775
 Kanamycin/neomycin resistance gene
 Neomycin phosphotransferase coding sequences:
 Start codon (ATG): 3858-3860; Stop codon: 4650-4652
 G->A mutation to remove Pst I site: 4040
 C->A (Arg to Ser) mutation to remove BssH II site: 4386
 Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal
 Polyadenylation signals: 4888-4893 & 4901-4906
 pUC plasmid replication origin: 5237-5880

References

Gorman, C. (1985). In: *DNA cloning: A Practical Approach*, Vol. II. Ed. by Glover. (IRL Press, Oxford, U.K.) Pp. 143-190.
 Haas, J. et al. (1996). *Curr Biol*, 6 (3): 315-324 / pmid: 8805248
 Kozak, M. (1987). *Nucleic Acids Res*, 15 (20): 8125-8148 / pmid: 3313277
 Li, X. et al. (1998). *J Biol Chem*, 273 (52): 34970-34975 / pmid: 9857028

Product	Cat.#	Size
peTurboGFP-dest1 vector	FP524	20 µg
Vector type	mammalian expression vector	
Reporter	TurboGFP	
Reporter codon usage	mammalian	
Promoter for TurboGFP	P _{CMV IE}	
Host cells	mammalian	
Selection	prokaryotic - kanamycin eukaryotic - neomycin (G418)	
Replication	prokaryotic - pUC ori eukaryotic - SV40 ori	
Use	TurboGFP expression in mammalian cells; generation of fusions to the TurboGFP-dest1 N-terminus	

Vector description

peTurboGFP-dest1 is a mammalian expression vector encoding destabilized variant of the green fluorescent protein TurboGFP. To generate TurboGFP-dest1 variant, residues 422-461 of mouse ornithine decarboxylase (MODC) were fused to the TurboGFP C-terminus. This MODC region contains a PEST amino acid sequence that targets the protein for degradation and provides for rapid protein turnover [Li et al. 1998]. TurboGFP-dest1 retains fluorescent properties of the native protein and has a half-life of approximately 1-1.5 hours, as measured by fluorescence intensity of cells treated with the protein synthesis inhibitor, cycloheximide.

peTurboGFP-dest1 carries synthetic version of the TurboGFP-dest1 gene which codon usage is optimized for high expression in mammalian cells (humanized) [Haas et al. 1996]. To increase mRNA translation efficiency, Kozak consensus translation initiation site is generated upstream of the TurboGFP-dest1 coding sequence [Kozak 1987]. Fragments of exons 2 and 3 and intron 2 of human beta globin gene are added in the 3' UTR of TurboGFP-dest1 coding sequence in order to increase the protein expression level.

peTurboGFP-dest1 vector can be used to express TurboGFP-dest1 in eukaryotic (mammalian) cells. For example it can be used as a positive control with a peTurboGFP-PRL-dest1 promoterless vector (Cat.# FP524). The vector can be also used to generate destabilized TurboGFP-tagged fusion proteins. Multiple cloning site (MCS) is located upstream of TurboGFP-dest1 coding sequence.

The vector backbone contains immediate early promoter of cytomegalovirus (P_{CMV IE}) for protein expression, SV40 origin for replication in mammalian cells expressing SV40 T-antigen, pUC origin of replication for propagation in *E. coli*, and f1 origin for single-stranded DNA production. SV40 polyadenylation signals (SV40 poly A) direct proper processing of the 3'-end of the reporter mRNA.

SV40 early promoter (P_{SV40}) provides neomycin resistance gene (Neo^r) expression to select stably transfected eukaryotic cells using G418. Bacterial promoter (P) provides kanamycin resistance gene expression (Kan^r) in *E. coli*. Kan^r/Neo^r gene is linked with herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signals.

Generation of TurboGFP-dest1-tagged fusions

A localization signal or a gene of interest can be cloned into MCS of the vector. It will be expressed as a fusion to the TurboGFP-dest1 N-terminus when inserted in the same reading frame as TurboGFP and no in-frame stop codons are present. TurboGFP-dest1-tagged fusions retain fluorescent properties of the native protein allowing fusion localization *in vivo*. Unmodified vector will express TurboGFP-dest1 when transfected into eukaryotic (mammalian) cells.

Note: The plasmid DNA was isolated from dam⁺-methylated *E. coli*. Therefore some restriction sites are blocked by methylation. If you wish to digest the vector using such sites you will need to transform the vector into a dam⁻ host and make fresh DNA.

Expression in mammalian cells

peTurboGFP-dest1 vector can be transfected into mammalian cells by any known transfection method. CMV promoter provides strong, constitutive expression of TurboGFP-dest1 or its fusions in eukaryotic cells. If required, stable transformants can be selected using G418 [Gorman 1985].

Propagation in *E. coli*

Suitable host strains for propagation in *E. coli* include DH5alpha, HB101, XL1-Blue, and other general purpose strains. Plasmid incompatibility group is pMB1/ColE1. The vector confers resistance to kanamycin (30 µg/ml) to *E. coli* hosts. Copy number in *E. coli* is about 500.

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