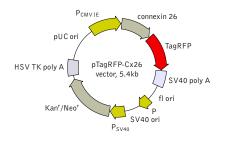


# pTagRFP-Cx26 vector

The vector sequence has been compiled using the information from sequence databases, published literature, and other sources, together with partial sequences obtained by Evrogen. This vector has not been completely sequenced.



For vector sequence, please visit our Web site at http://www.evrogen.com/products/vectors.shtml

## **Location of features**

P<sub>CMV IE</sub>: 1-589 Enhancer region: 59-465 TATA box: 554-560 Transcription start point: 583 Connexin 26: 683-1360 TagRFP: 1382-2095 SV40 early mRNA polyadenylation signal Polyadenylation signals: 2248-2253 & 2277-2282 mRNA 3' ends: 2286 & 2298 f1 single-strand DNA origin: 2345-2800 Bacterial promoter for expression of Kan<sup>r</sup> gene -35 region: 2862-2867; -10 region: 2885-2890 Transcription start point: 2897 SV40 origin of replication: 3141-3276 SV40 early promoter Enhancer (72-bp tandem repeats): 2974-3045 & 3046-3117 21-bp repeats: 3121-3141, 3142-3162 & 3164-3184 Early promoter element: 3197-3203 Major transcription start points: 3193, 3231, 3237 & 3242 Kanamycin/neomycin resistance gene Neomycin phosphotransferase coding sequences: Start codon (ATG): 3325-3327; Stop codon: 4117-4119

G->A mutation to remove Pst I site: 3507 C->A (Arg to Ser) mutation to remove BssH II site: 3853 Herpes simplex virus (HSV) thymidine kinase (TK)

polyadenylation signal Polyadenylation signals: 4355-4360 & 4368-4373

pUC plasmid replication origin: 4704-5347

Product	Cat.#	Size	
pTagRFP-Cx26 vector	FP362	20 $\mu$ g	
Vector type	mammalian expr	ression vector	
Reporter	TagRFP mammalian		
Reporter codon usage	mammalian		
Promoter for TagRFP	P <sub>CMV IE</sub>		
Host cells			
Selection	prokaryotic - kanamycin eukaryotic - neomycin (G418)		
Replication	prokaryotic - pUC ori eukaryotic - SV40 ori		
Use	red (orange) fluorescent labeling of connexin 26		

## Vector description

pTagRFP-Cx26 is a mammalian expression vector encoding TagRFP-Cx26 fusion protein. The vector can be used for fluorescent labeling of connexin 26 in living cells.

TagRFP codon usage is optimized for high expression in mammalian cells (humanized) [Haas et al. 1996]. Rat connexin 26 is fused to the TagRFP N-terminus.

pTagRFP-Cx26 vector can be used as a source of TagRFP-Cx26 hybrid sequence. The vector backbone contains unique restriction sites that permit its excision and further insertion into expression vector of choice. **Note:** The plasmid DNA was isolated from dam<sup>+</sup>-methylated *E.coli*. Therefore some restriction sites are blocked by methylation. If you wish to digest the vector using such sites you will need to transform the vector into a dam<sup>-</sup> host and make fresh DNA.

The vector backbone contains immediate early promoter of cytomegalovirus ( $P_{CMV \, IE}$ ) for protein expression, SV40 origin for replication in mammalian cells expressing SV40 T-antigen, pUC origin of replication for propagation in *E. coli*, and f1 origin for single-stranded DNA production. SV40 polyadenylation signals (SV40 poly A) direct proper processing of the 3'-end of the reporter mRNA.

SV40 early promoter (P<sub>SV40</sub>) provides neomycin resistance gene (Neo<sup>r</sup>) expression to select stably transfected eukaryotic cells using G418. Bacterial promoter (P) provides kanamycin resistance gene expression (Kan<sup>r</sup>) in *E. coli.* Kan<sup>r</sup>/Neo<sup>r</sup> gene is linked with herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signals.

## Expression in mammalian cells

pTagRFP-Cx26 vector can be transfected into mammalian cells by any known transfection method. CMV promoter provides strong, constitutive expression of the TagRFP-Cx26 fusion in eukaryotic cells. If required, stable transformants can be selected using G418 [Gorman 1985].

#### Propagation in E. coli

Suitable host strains for propagation in *E. coli* include DH5alpha, HB101, XL1-Blue, and other general purpose strains. Plasmid incompatibility group is pMB1/ColE1. The vector confers resistance to kanamycin (30  $\mu$ g/ml) to *E. coli* hosts. Copy number in *E. coli* is about 500.

#### References

Gorman, C. (1985). "High efficiency gene transfer into mammalian cells." In: DNA cloning: A Practical Approach, Vol. II. Ed. by Glover. (IRL Press, Oxford, U.K.) Pp. 143–190.

Haas, J. et al. (1996) "Codon usage limitation in the expression of HIV-1 envelope glycoprotein." Curr Biol, 6 (3): 315–324 / pmid: 8805248

#### Notice to Purchaser:

TagRFP-related materials (also referred to as "Products") are intended for research use only.

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MSDS information is available at http://www.evrogen.com/MSDS.shtml

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