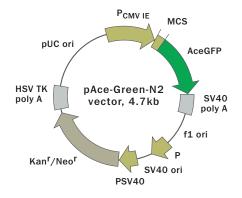


# Mammalian expression vector pAce-Green-N2



For vector sequence, please visit our Web site at www.evrogen.com/support/vector-info.shtml

Product	Cat.#	Size
pAce-Green-N2	FP105	20 μg

Please contact your local distributor for exact prices and delivery information.

Vector type mammalian expression vector

Reporter AceGFP
Reporter codon usage mammalian
Promoter for AceGFP
Host cells mammalian

Selection prokaryotic — kanamycin

eukaryotic — neomycin (G418)

AcaGED

Replication prokaryotic — pUC ori eukaryotic — SV40 ori

#### Multiple cloning site (MCS)

Nhel	BglII	Sacl	HindIII	EcoRI		Sall	Kpnl		Apal	BamHI	Agel			7000	<b>—</b>
GC. TAG. CGC. TAC. CGG.	ACT.CAG.ATC.TCG	G.AGC.TC	A. AGC. TTC	GAA.TTC.	TGC.A	GT. CGA.	CGG. TAC.	CGC.GG	G.CCC.	GGG. ATC. CAC	. CGG. CCG	.GTC.GCC	. ACC	ATG. A	IGC. AAG
Afel	Xho	ol .			Pstl	_	_	Sacll*	Smal/	/Xmal					

<sup>\* —</sup> not unique sites.

#### Use

# - Generation of fusions to the AceGFP N-terminus

# **Vector description**

pAce-Green-N2 is an eukaryotic (mammalian) expression vector encoding the *Aequrea coerulescens* enhanced green fluorescent protein AceGFP. The vector is designed to generate AceGFP-tagged fusions or to express AceGFP in mammalian cells.

AceGFP codon usage is optimized for high expression in mammalian cells (humanized, Haas et al., 1996). To increase AceGFP translation, Kozak consensus translation initiation site is generated upstream of AceGFP sequence (Kozak, 1987). Multiple cloning site (MCS) is located between  $P_{\text{CMV IE}}$  and AceGFP coding sequence.

The vector backbone comprises immediate early promoter of cytomegalovirus ( $P_{CMV \mid E}$ ) for protein expression, SV40 origin for replication in mammalian cells expressing SV40 T-antigen, pUC origin of replication for propagation in *E. coli* and f1 origin for single-stranded DNA production. SV40 polyadenylation signals (SV40 polyA) direct proper processing of the 3' end of the reporter mRNA.

SV40 early promoter provides neomycin resistance gene expression to select stably transfected eukaryotic cells using G418. Bacterial promoter (P) provides kanamycin resistance gene expression in *E. coli.* Kan<sup>r</sup>/Neo<sup>r</sup> gene is linked with herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signals.

### Generation of fusions

A localization signal (or a gene of interest) should be cloned into MCS of the vector. It will be expressed as a fusion to the AceGFP N-terminus when inserted in the same reading frame as AceGFP and no intervening stop codons are present. The inserted sequence should contain an initiating ATG codon. AceGFP fusions retain fluorescent properties of the native protein allowing fusion protein localization *in vivo*.

**Note:** The plasmid DNA was isolated from dam<sup>+</sup>-methylated *E.coli*. Therefore some restriction sites are bloked by methylation. If you wish to digest the vector using such sites you will need to transform the vector into a dam<sup>-</sup> host and make fresh DNA.

<sup>-</sup> Expression of AceGFP or its fusions in mammalian cells

### Expression in mammalian cells

pAce-Green-N2 can be transfected into mammalian cells by any known transfection method. If required, stable transformants can be selected using G418 (Gorman, 1985).

Unmodified pAce-Green-N1 will express AceGFP, when transfected into eukaryotic (mammalian) cells.

## Propagation in E. coli

Suitable host strains for propagation in E. coli include DH5alpha, HB101, XL1-Blue, and other general purpose strains. Plasmid incompatibility group is pMB1/CoIE1. The vector confers resistance to kanamycin (30 µg/ml) to E. coli hosts. Copy number in E. coli is about 500.

#### **Location of features**

P<sub>CMV IE</sub>: 1-589

Enhancer region: 59-465 TATA box: 554-560

Transcription start point: 583

MCS: 591-676 AceGFP

Kozak consensus translation initiation site: 677-687 Start codon (ATG): 679-681; Stop codon: 1401-1403

SV40 early mRNA polyadenylation signal

Polyadenylation signals: 1557-1563 & 1586-1591

mRNA 3' ends: 1595 & 1607

f1 single-strand DNA origin: 1654-2109 (packages the noncoding strand of AceGFP) Bacterial promoter expression of Kan<sup>r</sup> gene:

-35 region: 2171-2176; -10 region: 2194-2199 Transcription start point: 2206

SV40 origin of replication: 2450-2585

SV40 early promoter

Enhancer (72-bp tandem repeats): 2283-2354 & 2355-2426 21-bp repeats: 2430-2450, 2451-2471, & 2473-2493

Early promoter element: 2506-2512

Major transcription start points: 2502, 2540, 2546 & 2551

Kanamycin/neomycin resistance gene

Neomycin phosphotransferase coding sequences: Start codon (ATG): 2634-2636; stop codon: 3426-3428

G->A mutation to remove Pst I site: 2816

C->A (Arg to Ser) mutation to remove BssH II site: 3162

Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal

Polyadenylation signals: 3664-3669 & 3677-3682

pUC plasmid replication origin: 4013-4656

### References

Gorman C. (1985) In DNA cloning: A Practical Approach, Vol. II. Ed. D. M. Glover. (IRL Press, Oxford, U.K.), pp. 143-190.

Haas J. et al. (1996) Curr. Biol. 6: 315-324.

Kozak M. (1987) Nucleic Acids Res. 15:8125-8148.

### Notice to Purchaser:

This product contains a proprietary nucleic acid coding for a proprietary fluorescent protein(s) intended to be used by academic (non-commercial) entities and for research purposes only. Any use of the proprietary nucleic acid or protein other than for research use or by a commercial entity is strictly prohibited. Transfer of this product by purchaser to any other party is specifically prohibited.

The CMV promoter is covered under U.S. Patents 5,168,062 and 5,385,839 and its use is permitted for research purposes only. Any other use of the CMV promoter requires a license from the University of Iowa Research Foundation, 214 Technology Innovation Center, Iowa City, IA 52242.

# MATERIAL SAFETY DATA SHEET INFORMATION

To the best of our knowledge, these products do not require a Material Safety Data Sheet. However, all the properties of these products (and, if applicable, each of their components) have not been thoroughly investigated. Therefore, we recommend that you use gloves and eye protection, and wear a laboratory coat when working with these products.

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