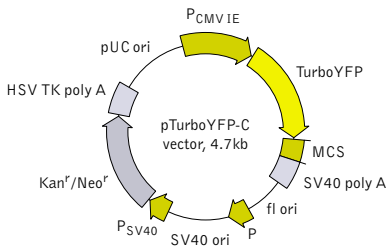


## pTurboYFP-C vector

The vector sequence has been compiled using the information from sequence databases, published literature, and other sources, together with partial sequences obtained by Evrogen. This vector has not been completely sequenced.



For vector sequence, please visit our Web site at <http://www.evrogen.com/support/vector-info.shtml>

### pTurboYFP-C vector MCS

TurboYFP → ... TCC. GGT. CTC. AGA. TCT. CGA. GCT. CAA. GCT. TCG. AAT. TCT. GCA. GTC. GAC. GGT. ACC. GCG. GGC. CCG. GGA. TCC. ACC. GGA. TCT. AGA. TAA. CTG. ATC. A . . .

*Xho I*
*Hind III*
*Pst I*
*Kpn I*
*Apa I*
*BamH I*
*STOPIs*

*Bgl II*
*Sac I*
*EcoRI*
*Sal I*
*Sac II\**
*Sma I/Xma I*
*Xba I#*
*Bcl I#*

\* — not unique site.

# — sites are blocked by *dam* methylation. If you wish to digest the vector with these enzymes, you will need to transform the vector into a *dam*<sup>-</sup> host and make fresh DNA.

### Location of features

P<sub>CMV IE</sub>: 1-589  
 Enhancer region: 59-465  
 TATA box: 554-560  
 Transcription start point: 583  
 TurboYFP  
 Kozak consensus translation initiation site: 606-616  
 Start codon (ATG): 613-615; Stop codon: 1420-1422  
 Last amino acid in TurboYFP: 1339-1341  
 MCS: 1342-1419  
 SV40 early mRNA polyadenylation signal  
 Polyadenylation signals: 1562-1567 & 1591-1596  
 mRNA 3' ends: 1600 & 1612  
 f1 single-strand DNA origin: 1659-2114  
 Eukaryotic promoter for expression of Kan<sup>r</sup> gene  
 -35 region: 2176-2181; -10 region: 2199-2204  
 Transcription start point: 2211  
 SV40 origin of replication: 2455-2590  
 SV40 early promoter  
 Enhancer (72-bp tandem repeats): 2288-2359 & 2360-2431  
 21-bp repeats: 2435-2455, 2456-2476 & 2478-2498  
 Early promoter element: 2511-2517  
 Major transcription start points: 2507, 2545, 2551 & 2556  
 Kanamycin/neomycin resistance gene  
 Neomycin phosphotransferase coding sequences:  
 Start codon (ATG): 2639-2641; Stop codon: 3431-3433  
 G->A mutation to remove Pst I site: 2821  
 C->A (Arg to Ser) mutation to remove BssH II site: 3167  
 Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal  
 Polyadenylation signals: 3669-3674 & 3682-3687  
 pUC plasmid replication origin: 4018-4661

### References

Gorman (1985). "High efficiency gene transfer into mammalian cells." In: *DNA cloning: A Practical Approach, Vol. II*. Ed. by Glover. (IRL Press, Oxford, U.K.) Pp. 143-90.

Haas et al. (1996) "Codon usage limitation in the expression of HIV-1 envelope glycoprotein." *Curr Biol*, 6 (3): 315-24 / pmid: 8805248

Kozak (1987) "An analysis of 5'-noncoding sequences from 699 vertebrate messenger RNAs." *Nucleic Acids Res*, 15 (20): 8125-48 / pmid: 3313277

Product	Cat.#	Size
pTurboYFP-C vector	FP611	20 µg
The price does not include delivery. The price varies in different countries. Please contact your local distributor for exact prices and delivery information.		
Vector type	mammalian expression vector	
Reporter	TurboYFP	
Reporter codon usage	mammalian	
Promoter for TurboYFP	P <sub>CMV IE</sub>	
Host cells	mammalian	
Selection	prokaryotic - kanamycin eukaryotic - neomycin (G418)	
Replication	prokaryotic - pUC ori eukaryotic - SV40 ori	
Use	TurboYFP expression in mammalian cells; generation of fusions to the TurboYFP C-terminus	

### Vector description

pTurboYFP-C is a mammalian expression vector encoding yellow fluorescent protein TurboYFP. The vector allows generation of fusions to the TurboYFP C-terminus and expression of TurboYFP fusions or TurboYFP alone in eukaryotic (mammalian) cells.

TurboYFP codon usage is optimized for high expression in mammalian cells (humanized) [Haas et al. 1996]. To increase mRNA translation efficiency, Kozak consensus translation initiation site is generated upstream of the TurboYFP sequence [Kozak 1987]. Multiple cloning site (MCS) is located between TurboYFP coding sequence and SV40 polyadenylation signal (SV40 polyA).

The vector backbone contains immediate early promoter of cytomegalovirus (P<sub>CMV IE</sub>) for protein expression, SV40 origin for replication in mammalian cells expressing SV40 T-antigen, pUC origin of replication for propagation in *E. coli* and f1 origin for single-stranded DNA production. SV40 polyadenylation signals (SV40 polyA) direct proper processing of the 3'-end of the reporter mRNA.

SV40 early promoter (P<sub>SV40</sub>) provides neomycin resistance gene (Neo<sup>r</sup>) expression to select stably transfected eukaryotic cells using G418. Bacterial promoter (P) provides kanamycin resistance gene expression (Kan<sup>r</sup>) in *E. coli*. Kan<sup>r</sup>/Neo<sup>r</sup> gene is linked with herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signals.

### Generation of TurboYFP-fusion proteins

A localization signal (or a gene of interest) should be cloned into MCS of the vector. It will be expressed as a fusion to the TurboYFP C-terminus when inserted in the same reading frame as TurboYFP and no intervening stop codons are present. TurboYFP-tagged fusions retain fluorescent properties of the native protein allowing fusion localization *in vivo*. Unmodified vector will express TurboYFP, when transfected into eukaryotic (mammalian) cells.

**Note:** The plasmid DNA was isolated from *dam*<sup>+</sup>-methylated *E. coli*. Therefore some restriction sites are blocked by methylation. If you wish to digest the vector using such sites you will need to transform the vector into a *dam*<sup>-</sup> host and make fresh DNA.

Despite its dimeric structure, TurboYFP is still suitable for generation of fusions with proteins of interest, however we recommend to use TagFPs for these purposes.

### Expression in mammalian cells

pTurboYFP-C vector can be transfected into mammalian cells by any known transfection method. If required, stable transformants can be selected using G418 [Gorman 1985].

### Propagation in *E. coli*

Suitable host strains for propagation in *E. coli* include DH5alpha, HB101, XL1-Blue, and other general purpose strains. Plasmid incompatibility group is pMB1/ColE1. The vector confers resistance to kanamycin (30 µg/ml) to *E. coli* hosts. Copy number in *E. coli* is about 500.

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