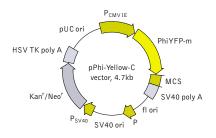


# pPhi-Yellow-C vector

The vector sequence has been compiled using the informa-tion from sequence databases, published literature, and other sources, together with partial sequences obtained by Evrogen. This vector has not been completely sequenced.



or vector sequence, please visit our Web site at

Product	Cat.#	Size							
pPhi-Yellow-C vector	FP601	$20~\mu \mathrm{g}$							
Vector type	mammalian expr	ession vector							
Reporter	PhiYFP-m								
Reporter codon usage	mammalian								
Promoter for PhiYFP-m	P <sub>CMV IE</sub>								
Host cells	mammalian								
Selection	prokaryotic - kana eukaryotic - neon	•							
Replication	prokaryotic - pUC eukaryotic - SV40								
Use	PhiYFP-m expression in mammalian cells; gen fusions to the PhiYFP-m C-terminus								

#### Multiple cloning site (MCS)

		Sac	I		Е	coR I		Sa	1 I			Sac II	* S	ma I/X	Kma I				Xb	a I#		В	cl I#	
PhiYFP-m	X	ho I	H	ind III		Щ_	Pst 1			Κį	on I		Apa —	I	Ba	mH I					ST0Ps			
GGA.	TCT. C	GA. GCT	. CAA.	GCT.	TCG. A	AT.TCT	. GCA	GTC.	GAC	GGT.	ACC.	. GCG.	GGC.	CCG.	GGA.	TCC.	ACC.	GGA.	TCT.	AGA.	TAA.	CTG.	ATC.A.	
G	S	R A	Ω	Α	S I	N S	Α	V	D	G	Т	Α	G	Р	G	S	Т	G	S	R	*	1	T	

# **Location of features**

P<sub>CMV IE</sub>: 1-589 Enhancer region: 59-465

TATA box: 554-560

Transcription start point: 583 PhiYFP-m

Kozak consensus translation initiation site: 606-616 Start codon (ATG): 613-615; Stop codon: 1384-1386 Last amino acid in PhiYFP-m: 1312-1314

MCS: 1315-1400

SV40 early mRNA polyadenylation signal Polyadenylation signals: 1526-1531 & 1555-1560

mRNA 3' ends: 1564 & 1576

f1 single-strand DNA origin: 1623-2078

Bacterial promoter for expression of Kan<sup>r</sup> gene -35 region: 2140-2145; -10 region: 2163-2168

Transcription start point: 2175

SV40 origin of replication: 2419-2554

SV40 early promoter

Enhancer (72-bp tandem repeats): 2252-2323 & 2324-

2395

21-bp repeats: 2399-2419, 2420-2440 & 2442-2462

Early promoter element: 2475-2481

Major transcription start points: 2471, 2509, 2515 & 2520

Kanamycin/neomycin resistance gene

Neomycin phosphotransferase coding sequences: Start codon (ATG): 2603-2605; Stop codon: 3395-3397 G->A mutation to remove Pst I site: 2785

C->A (Arg to Ser) mutation to remove BssH II site: 3131 Herpes simplex virus (HSV) thymidine kinase (TK)

polyadenylation signal Polyadenylation signals: 3633-3638 & 3646-3651

pUC plasmid replication origin: 3982-4625

#### References

Gorman, C. (1985), "High efficiency gene transfer into mammalian cells." In: DNA cloning: A Practical Approach, Vol. II. Ed. by Glover. (IRL Press, Oxford, U.K.)

Haas, J. et al. (1996) "Codon usage limitation in the expression of HIV-1 envelope glycoprotein." Curr Biol, 6 (3): 315-324 / pmid: 8805248

Kozak, M. (1987) "An analysis of 5'-noncoding sequences from 699 vertebrate messenger RNAs." Nucleic Acids Res, 15 (20): 8125-8148 / pmid: 3313277

# **Vector description**

pPhi-Yellow-C is a mammalian expression vector encoding yellow fluorescent protein PhiYFP-m. The vector allows generation of fusions to the PhiYFP-m C-terminus and expression of PhiYFP-m fusions or PhiYFP-m alone in eukaryotic (mammalian) cells.

PhiYFP-m codon usage is optimized for high expression in mammalian cells (humanized) [Haas et al. 1996]. To increase mRNA translation efficiency, Kozak consensus translation initiation site is generated upstream of the PhiYFP-m coding sequence [Kozak 1987]. Multiple cloning site (MCS) is located between PhiYFP-m coding sequence and SV40 polyadenylation signal (SV40 polyA).

The vector backbone contains immediate early promoter of cytomegalovirus (P<sub>CMV IE</sub>) for protein expression, SV40 origin for replication in mammalian cells expressing SV40 T-antigen, pUC origin of replication for propagation in E. coli, and f1 origin for single-stranded DNA production. SV40 polyadenylation signals (SV40 poly A) direct proper processing of the 3'-end of the reporter mRNA.

SV40 early promoter (P<sub>SV40</sub>) provides neomycin resistance gene (Neo<sup>r</sup>) expression to select stably transfected eukaryotic cells using G418. Bacterial promoter (P) provides kanamycin resistance gene expression (Kan<sup>r</sup>) in E. coli. Kan<sup>r</sup>/Neo<sup>r</sup> gene is linked with herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signals.

# **Generation of PhiYFP-m fusion proteins**

A localization signal or a gene of interest can be cloned into MCS of the vector. It will be expressed as a fusion to the PhiYFP-m C-terminus when inserted in the same reading frame as PhiYFP-m and no in-frame stop codons are present. PhiYFP-m-tagged fusions retain fluorescent properties of the native protein allowing fusion localization in vivo. Unmodified vector will express PhiYFP-m when transfected into eukaryotic (mammalian) cells.

Note: The plasmid DNA was isolated from dam +-methylated E.coli. Therefore some restriction sites are blocked by methylation. If you wish to digest the vector using such sites you will need to transform the vector into a dam host and make fresh DNA.

#### Expression in mammalian cells

pPhi-Yellow-C vector can be transfected into mammalian cells by any known transfection method. CMV promoter provides strong, constitutive expression of PhiYFP-m or its fusions in eukaryotic cells. If required, stable transformants can be selected using G418 [Gorman 1985].

# Propagation in E. coli

Suitable host strains for propagation in E. coli include DH5alpha, HB101, XL1-Blue, and other general purpose strains. Plasmid incompatibility group is pMB1/CoIE1. The vector confers resistance to kanamycin (30  $\mu$ g/ml) to E. coli hosts. Copy number in E. coli is about 500.

#### **Notice to Purchaser:**

PhiYFP-related materials (also referred to as "Products") are intended for research use only.

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The CMV promoter is covered under U.S. Patents 5,168,062 and 5,385,839, and its use is permitted for research purposes only. Any other use of the CMV promoter requires a license from the University of Iowa Research Foundation, 214 Technology Innovation Center, Iowa City, IA 52242

<sup>-</sup> sites are blocked by dam methylation. If you wish to digest the vector with these enzymes, you will need to transform the vector into a dam host and make fresh DNA.