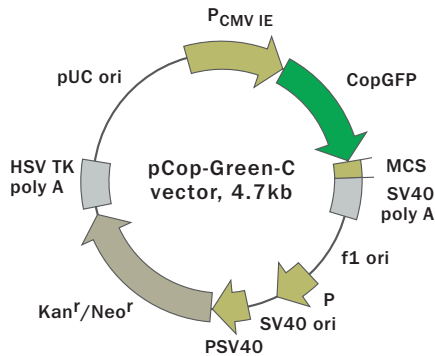


Mammalian expression vector pCop-Green-C



For vector sequence, please visit our Web site at www.evrogen.com/support/vector-info.shtml

| Product | Cat.# | Size |
|--------------|-------|-------|
| pCop-Green-C | FP501 | 20 µg |

Please contact your local distributor for exact prices and delivery information.

| | |
|----------------------|---|
| Vector type | mammalian expression vector |
| Reporter | CopGFP |
| Reporter codon usage | mammalian |
| Promoter for CopGFP | P _{CMV IE} |
| Host cells | mammalian |
| Selection | prokaryotic — kanamycin eukaryotic — neomycin (G418) |
| Replication | prokaryotic — pUC ori eukaryotic — SV40 ori |

Multiple cloning site (MCS)



* - not unique sites. # - sites are blocked by methylation.

Use

- Generation of fusions to the CopGFP C-terminus
- Expression of CopGFP or its fusions in mammalian cells

Vector description

pCop-Green-C is an eukaryotic (mammalian) expression vector encoding green fluorescent protein CopGFP. The vector allows generation of fusions to the CopGFP C-terminus and expression CopGFP fusions or CopGFP alone in eukaryotic (mammalian) cells.

CopGFP codon usage is optimized for high expression in mammalian cells (humanized, Haas *et al.*, 1996). To increase CopGFP translation, Kozak consensus translation initiation site is generated upstream of the CopGFP sequence (Kozak, 1987). Multiple cloning site (MCS) is located between CopGFP coding sequence and SV40 polyadenylation signal (SV40 polyA).

The vector backbone contains immediate early promoter of cytomegalovirus (P_{CMV IE}) for protein expression, SV40 origin for replication in mammalian cells expressing SV40 T-antigen, pUC origin of replication for propagation in *E. coli*, and f1 origin for single-stranded DNA production. SV40 polyA direct proper processing of the 3' end of the reporter mRNA.

SV40 early promoter provides neomycin resistance gene expression to select stably transfected eukaryotic cells using G418. Bacterial promoter (P) provides kanamycin resistance gene expression in *E. coli*. Kan^r/Neo^r gene is linked with herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signals.

Generation of fusions

A localization signal (or a gene of interest) should be cloned into MCS of the vector. It will be expressed as a fusion to CopGFP C-terminus when inserted in the same reading frame as CopGFP and no intervening stop codons are present. CopGFP-tagged fusions retain fluorescent properties of the native protein allowing fusion localization *in vivo*. Unmodified vector will express CopGFP, when transfected into eukaryotic (mammalian) cells.

Note: The plasmid DNA was isolated from dam⁺-methylated *E. coli*. Therefore some restriction sites are blocked by methylation. If you wish to digest the vector using such sites you will need to transform the vector into a dam⁻ host and make fresh DNA.

Expression in mammalian cells

pCop-Green-C can be transfected into mammalian cells by any known transfection method. If required, stable transformants can be selected using G418 (Gorman, 1985).

Propagation in *E. coli*

Suitable host strains for propagation in *E. coli* include DH5alpha, HB101, XL1-Blue, and other general purpose strains. Plasmid incompatibility group is pMB1/ColE1. The vector confers resistance to kanamycin (30 µg/ml) to *E. coli* hosts. Copy number in *E. coli* is about 500.

Location of features

P_{CMV IE}: 1-589

Enhancer region: 59-465

TATA box: 554-560

Transcription start point: 583

CopGFP

Kozak consensus translation initiation site: 606-616

Start codon (ATG): 613-615;

Stop codon: 1348-1350

Last amino acid in CopGFP: 1376-1378

MCS: 1279-1357

SV40 early mRNA polyadenylation signal

Polyadenylation signals: 1490-1495 & 1519-1524

mRNA 3' ends: 1528 & 1540

f1 single-strand DNA origin: 1587-2042

(packages the noncoding strand of CopGFP)

Bacterial promoter for expression of Kan^r gene

-35 region: 2160-2165;

-10 region: 2127-2132

Transcription start point: 2139

SV40 origin of replication: 2383-2518

SV40 early promoter

Enhancer (72-bp tandem repeats): 2216-2287 & 2288-2359

21-bp repeats: 2363-2383, 2384-2404, & 2406-2426

Early promoter element: 2439-2445

Major transcription start points: 2435, 2473, 2479 & 2484

Kanamycin/neomycin resistance gene

Neomycin phosphotransferase coding sequences:

Start codon (ATG): 2567-2569; stop codon: 3359-3361

G->A mutation to remove PstI site: 2749

C->A (Arg to Ser) mutation to remove BssH II site: 3095

Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal

Polyadenylation signals: 3597-3602 & 3610-3615

pUC plasmid replication origin: 3946-4589

References

Gorman C. (1985) In DNA cloning: A Practical Approach, Vol. II. Ed. D. M. Glover. (IRL Press, Oxford, U.K.), pp. 143-190.

Haas J. *et al.* (1996) *Curr. Biol.* 6: 315-324.

Kozak M. (1987) *Nucleic Acids Res.* 15:8125-8148.

Notice to Purchaser:

Evrogen Fluorescent Protein Products (the Products) are intended for research use only and covered by Evrogen Patents and/or Patent applications pending. By use of these products, you accept the terms and conditions of the applicable Limited Use Label License (enclosed).

CMV promoter: The CMV promoter is covered under U.S. Patents 5,168,062 and 5,385,839 and its use is permitted for research purposes only. Any other use of the CMV promoter requires a license from the University of Iowa Research Foundation, 214 Technology Innovation Center, Iowa City, IA 52242.

MATERIAL SAFETY DATA SHEET INFORMATION:

To the best of our knowledge, these products do not require a Material Safety Data Sheet. However, all the properties of these products (and, if applicable, each of their components) have not been thoroughly investigated. Therefore, we recommend that you use gloves and eye protection, and wear a laboratory coat when working with these products.